A Case Series of Percutaneous Treatment of Non-Union Fractures with Autologous, Culture Expanded, Bone Marrow Derived, Mesenchymal Stem Cells and Platelet Lysate

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Abstract

Background: Current treatment options for stable non-union fractures represent major clinical challenges, and are a major health issue. Fracture treatment can take many forms, usually requiring bone grafting and/or revisions of the fracture with open reduction and internal fixation (ORIF). Conservative care options such as bone morphogenic proteins and bone stimulators are also available. The purpose of this study was to determine if culture expanded, autologous MSC’s injected into non-union fractures under c-Arm fluoroscopy could represent an alternative treatment modality in recalcitrant fracture non-unions. This paper reports on the findings of 6 patients with fracture non-union treated with autologous MSC’s.

Patients and methods: We evaluated 6 consecutive patients with chronic fracture non-unions. Patients consisted of 4 women and 2 men with treatment intervention at an average of 8.7 months post-fracture (range 4-18 months, one patient fracture not included in calculation was >100 mo.). All treated patients received autologous, culture expanded, mesenchymal stem cells injected percutaneously via fluoroscopic guidance into the site of the fracture non-union. Fracture union was evaluated with the use of follow up high-resolution x-ray and/or CT imaging. Phenotype of the culture-expanded MSC’s was evaluated and quantified by flow cytometry of surface antigens.

Conclusion: The results of this study support the hypothesis that autologous MSC’s delivered via percutaneous re-implantation may be an alternative modality for the non-operative treatment of recalcitrant non-union fractures.

Introduction

The economic burden of non-union fractures is significant due to the cost and disability associated with the condition. Of the approximate 5.6 million bone fractures in the United States annually, up to 10% do not fully heal, and require subsequent treatment [1]. The traditional treatment options for fracture non-union run the spectrum from less invasive bone stimulation techniques using ultrasound and pulsed electromagnetic fields to more invasive interventions including autologous or allogeneic bone grafting either with or without the aid of internal/external fixation. Osteobiologics offer yet another existing modality in recalcitrant fracture non-unions. This paper reports on the findings of 6 patients with fracture non-union treated with autologous MSC’s.

Fracture healing is a multidimensional process consisting of four well established remodeling stages; an initial inflammatory response, soft callus formation, initial bony union and bone remodeling. At the cellular level, inflammatory cells, vascular cells, osteochondral progenitors including mesenchymal stem cells (MSC’s), and osteoclasts are fundamental in the repair process [4]. MSC’s are pluripotent cells found in multiple human adult tissues including bone marrow, synovial tissues, and adipose tissues. Since they are derived from the mesoderm, they have the capacity to differentiate into bone, cartilage, muscle, and adipose tissue [5]. Iwakura et al. [6] demonstrated the hematoma at the initial fracture site contains multi-lineage mesenchymal progenitor cells which are critical in the role in bone healing. Animal and early clinical models have shown that MSC’s can be used to regenerate articular cartilage and bone [7-20].

Hernigou has used bone marrow selection to concentrate MSC’s and hematopoietic stem cells. This concentrate is then surgically re-implanted into fracture lines to assist in the healing of non-unions [2]. Muschler et al. [21] have also demonstrated a system that relies on selective retention of heterogeneous bone marrow nucleated cells onto a demineralized bone graft. This graft is then re-implanted surgically to promote fusion. A randomized trial of percutaneously injected cultured osteoblasts to treat fractures has been published by Kim et al. [22] showing acceleration in the rate of normal fracture healing. We have previously published on the safety of cultured MSC’s for clinical use as well as on selected cases showing orthopedic tissue regeneration [23-25]. However, at the time of the submission of this manuscript, we could find no published work on the use of percutaneously implanted, cultured MSC’s as a treatment for fracture non-union. There are obvious advantages to a model that relies on percutaneous injection, as this delivery method would dramatically reduce morbidity over open surgical implant methods.

The purpose of this study is to investigate the feasibility of a less invasive, percutaneous approach for the reimplantation of autologous culture expanded, bone marrow derived MSC’s in platelet lysate to enhance fracture repair in recalcitrant stable non-union cases.

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Material and methods

The research protocol was approved through a non-profit Institutional Review Board (Spinal Injury Foundation-IRB00002637). The definition of fracture non-union differs based on author from 3 consecutive months without radiographic evidence of healing to a minimum of 9 months without union, depending on site and whether infection was involved [26-29]. For the purposes of this investigation, we considered a non-union to be three months without evidence of bridging osteophytes.

Inclusion criteria

Male or female patients, 18-65 years of age
1. Radiographic evidence of disabling fracture non-union or delayed union with no progression of fracture healing over three consecutive months. Non progression of fracture healing was further defined on radiographs by:
   a. absence of bone crossing the fracture site
   b. persistent fracture line(s)
   c. lack of evidence of progression of healing on serial imaging

Exclusion criteria:

1. Active inflammatory or connective tissue disease (i.e. lupus, RA)
2. Active non-corrected endocrine disorder potentially associated with symptoms (i.e. hypothyroidism, diabetes)
3. Severe cardiac disease
4. Pulmonary disease requiring medication usage

Pre-procedure data collection:

Peripheral blood count (CBC) and a multiple serum panel (SMAC) to rule out unknown medical condition (within 3 months of procedure)

Re-implant procedure

In order to accurately place stem cells into the fracture line, fluoroscopy with injection of radiographic contrast confirming cannulation of the fracture line was utilized. Omnipaque (Iohexol-GE Healthcare NDC 0407-1413-50) is a non-ionic contrast agent that was used in the present study after first being diluted (1:1 or 1:2) with phosphate buffered saline. We have previously published on the effects of Iohexol on MSC’s, demonstrating minimal effects on cell viability [25].

Full informed consent was used prior to the procedure. For one week prior to the marrow harvest procedure the patient was restricted from taking corticosteroids or NSAIDs. Coincident with the marrow harvest procedure, approximately 200cc of heparinized IV venous blood was drawn to be used for platelet lysate (PL). Platelet lysate was prepared via centrifugation at 200g to separate platelet rich plasma (PRP) from the red blood cells (RBCs). The PRP was drawn off and stored at -20°C to produce PL. Platelet lysate was supplemented in cell culture media at 10-20% v/v.

For the marrow aspiration procedure, the patient was then placed prone on an operating room (OR) table and the area targeted for MSC harvest (Figure 1) was anesthetized with 1% Lidocaine, and a sterile disposable trocar was used to draw 10 cc of marrow blood from the right posterior superior iliac spine (PSIS), and 10cc from the left PSIS in syringes containing 1,000 U. of Heparin per ml (Abraxis Pharmaceuticals). MSC processing and culture expansion techniques have been described in prior publications [25]. Briefly, nucleated cells were isolated via centrifugation and plated in monolayer culture using autologous platelet lysate. After MSC’s had been grown to the 3rd-7th culture passage, they were suspended in phosphate buffered saline (PBS) and platelet lysate. Cell characterization and viability determination was via microscopic morphology grading and intermittent flow cytometry to ensure an MSC phenotype [25].

For MSC re-implantation, the patient was returned to the OR and the area of the non-union site was prepped using betadine and sterile gloves. A sterile trocar (11 gauge x 4-Tyco Healthcare-8881247111) was then inserted using c-arm fluoroscopy as guidance (Seimens Iso-C) into the visible fracture site at several locations. At each site, Omnipaque (300mg/ml) contrast was injected and once proper dye flow was demonstrated, cells were injected in a slow and controlled fashion. The trocar was then extracted and the patient was kept in the same position for one hour to allow for cell attachment. Post procedure

<table>
<thead>
<tr>
<th>ID</th>
<th>Stem Cell Yield at Injection ( Millions)</th>
<th>Passage at Injection</th>
<th>Days in Culture at Injection</th>
</tr>
</thead>
<tbody>
<tr>
<td>SB</td>
<td>29.92</td>
<td>5</td>
<td>22</td>
</tr>
<tr>
<td>JA</td>
<td>22.32</td>
<td>7</td>
<td>25</td>
</tr>
<tr>
<td>SM</td>
<td>5</td>
<td>5</td>
<td>21</td>
</tr>
<tr>
<td>MO</td>
<td>96</td>
<td>5</td>
<td>24</td>
</tr>
<tr>
<td>EW</td>
<td>3.15</td>
<td>2</td>
<td>17</td>
</tr>
<tr>
<td>BS</td>
<td>25.1</td>
<td>3</td>
<td>18</td>
</tr>
<tr>
<td>Average</td>
<td>30.25</td>
<td>4.5</td>
<td>21.2</td>
</tr>
<tr>
<td>St Dev</td>
<td>34.01</td>
<td>1.76</td>
<td>3.19</td>
</tr>
</tbody>
</table>

Table 1: Cell culture results.
discharge instructions included returning to activity as tolerated. Post procedure images were obtained at approximately 1 month, 3 months, and if needed at 6+ months, using matching views (when available).

Results

Six consecutive fracture patients were actively recruited by an office based, interventional pain practice based on diagnosis and their willingness to proceed with the study. This convenience sample consisted of 6 stable fracture non-unions at various sites. Average time in culture was 21.2 days ± 3.19 with average MSCs injected at 30.25 X 10⁶ ± 34.01. The average passage at injection was the 4th passage (4.5) with a standard deviation of ± 1.8 passages (Table 1). Cell morphology in culture was spindle shaped, typical of MSCs in monolayer culture and intermittent flow cytometry of cells grown in culture with this technique demonstrated cell surface antigens consistent with MSC’s. (Figure 2).

Of the 6 consecutive patients treated, adequate healing and return to functional recovery was seen in four patients (one patient lost to follow-up-see Table 2). The only fracture that did not heal was the oldest fracture (40 years duration).

Individual Patient Results

SB was a 37 year old WF with a 9 month old traumatic, non-displaced, fracture non-union of the distal humerus that had failed ORIF and a bone stimulator trial of 9 months duration (DonJoy CMF OL1000 Bone Growth Stimulator, DJO Inc.; Vista, CA USA). Co-morbidities included a ¼ pack per day smoking history with a total of 8 pack-years and morbid obesity. Pre-op radiographs showed no healing (see Figure 3a). Post-treatment radiographs at 5 weeks showed good early healing of the distal humerus fracture (see Figure 3b). The patient returned to full activities by the 2nd month following implant of MSC’s and demonstrated interval healing at 4 months. Final radiograph at 14 months showed excellent healing (Figure 3c).

SB fracture of distal humerus at 9 months post ORIF and after the failure of a bone stimulator.

SM was a 68 year old WF with a long history of multiple sclerosis and significant functional limitation in ambulation related to mild hemiparesis. She presented with an 18 month old fracture non-union of the 1st metatarsal that had failed 18 months of immobilization in a walking boot immobilizer. It should be noted that the patient complained of pain during the percutaneous MSC transplant procedure and as a result, she was unable to tolerate complete insertion of a 22 gauge needle into the faintly visible fracture line (a trocar was deemed too large for this fracture site). Cells were thus deployed of a 22 gauge needle into the faintly visible fracture line (a trocar was deemed too large for this fracture site). Follow-up thin slice CT demonstrated “moderate changes of healing across the superior one-half of both right and left sacral insufficiency fractures” as read by the reading radiologist (site treated). Eight month post-transplant thin slice CT demonstrated “advanced healing across bilateral sacral insufficiency fractures” as read by the same reading radiologist. From a functional perspective, within 4-6 weeks of the transplant, the patient was fully ambulatory with only age appropriate functional limitations.

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MO was 59 year old WF with a 40 year history of traumatic ischial tuberosity avulsion fracture and proximal hamstring tear. She had a long history of chronic low back pain managed with medications and epidural steroid injections. Two attempts were made at treating the fracture site since she complained of chronic pain localized to the ischial tuberosity which was responsive to local anesthetic injection. Post-treatment thin slice CT scans after both treatments failed to yield any signs of healing as read by the reading radiologist.

EW was a 67 year old WM s/p ORIF with a 4 month old traumatic, tibia-fibula non-union (figure 4a). The patient had difficulty with ambulation. Co-morbidities included chronic low back pain with high-risk opioid pain management, type II diabetes mellitus, and coronary artery disease. An MSC transplant was undertaken for the tibia and fibular fractures. While MSC’s could be easily placed into the tibial fracture line, the patient reported unacceptable levels of peroneal distribution pain with attempted cannulation of the fibular fracture, so that transplant site was aborted without MSC transplantation. The patient reported return to full ambulation with significant improvements in pain and function at 4-6 weeks post-transplant.

Table 2: Summary of patient demographics and treatment results.

<table>
<thead>
<tr>
<th>ID</th>
<th>Sex</th>
<th>Age</th>
<th>Co-morbidities</th>
<th>Duration of non-union (Mo)</th>
<th>Site Treated</th>
<th>Conservative Care</th>
<th>Image Timing (Mo)</th>
<th>Imaging</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>SB</td>
<td>F</td>
<td>37</td>
<td>Smoking, Morbid Obesity</td>
<td>9</td>
<td>Humerus</td>
<td>Immobilization, Bone Stimulator</td>
<td>Pre, 1, 2, 4, 14</td>
<td>Radiograph</td>
<td>Complete healing</td>
</tr>
<tr>
<td>JA</td>
<td>F</td>
<td>82</td>
<td>Severe Osteoporosis</td>
<td>4</td>
<td>Sacral Base</td>
<td>PT, Immobilization</td>
<td>Pre, 2, 8</td>
<td>Thin Slice CT</td>
<td>Advanced healing</td>
</tr>
<tr>
<td>SM</td>
<td>F</td>
<td>68</td>
<td>Multiple Sclerosis</td>
<td>18</td>
<td>1st meta-tarsal</td>
<td>Immobilization</td>
<td>Pre, 2, 8</td>
<td>Thin Slice CT</td>
<td>Moderate healing</td>
</tr>
<tr>
<td>MO</td>
<td>F</td>
<td>59</td>
<td>Chronic Low Back Pain</td>
<td>&gt;100</td>
<td>Ischial Tuberosity</td>
<td>None</td>
<td>Pre, 1.5, 3</td>
<td>Thin Slice CT</td>
<td>No healing detected</td>
</tr>
<tr>
<td>EW</td>
<td>M</td>
<td>67</td>
<td>Type II DM, CAD, Chronic Low Back Pain</td>
<td>4</td>
<td>Tibia</td>
<td>None</td>
<td>Pre, 2, 5</td>
<td>Radiograph</td>
<td>Advanced healing</td>
</tr>
<tr>
<td>BS</td>
<td>M</td>
<td>25</td>
<td>Peroneal nerve injury</td>
<td>24</td>
<td>Tibia-Fibula</td>
<td>PT, Immobilization</td>
<td>Pre, lost to f/u</td>
<td>Radiograph</td>
<td>Unknown</td>
</tr>
</tbody>
</table>

Figure 4b shows good healing of the tibial fracture site at 5 months post-transplant. More frequent interval images on this patient proved difficult as his work frequently prevented him from maintaining follow up.

BS is a 25 year old male status post traumatic tibia-fibula fracture with intramedulary rodding without complete healing at two years' post rodding. The patient had not returned to full activities and pain was 5/10. Comorbidities included a likely peroneal nerve injury at that site, detected on initial exam, but not confirmed on EMG/NCS. The tibia fracture was treated by MSC re-implantation into the site. The patient was lost to follow-up, despite multiple attempts to contact him for follow-up data and radiographs.

Discussion

Four of the six areas treated with percutaneously implanted MSC’s showed significant healing on radiographs and good return to full pre-fracture functional activities. The only fracture that showed no signs of healing was a fracture of 40 years duration that was difficult to immobilize (all other patients had immobilization with ORIF or didn’t require immobilization due to the nature of the fracture). It is unknown if this treatment failure was due to fracture duration, the nature of the fracture (avulsion fracture), the likely atrophic nature of the local bone healing environment, or the lack of immobilization. Bone reacts to its strain environment to provide signals for cellular healing mechanisms [30]. It is certainly possible that the strain environment of this well adapted, old fracture could no longer produce signals for cellular repair.

JA presented an interesting treatment dilemma. Unlike some senile fractures due to osteoporosis, our analysis demonstrated that stabilizing her non-healing sacral base fracture with injection of PMMA cement would be risky, as communication of the fracture line into the S2 foramen could have led to inadvertent nerve root compression. It should also be noted that JA’s outcome was surprising, given that she demonstrated severe osteoporosis both on radiographs and the tactile feel of the trocar at time of treatment. Arguments in the published literature on whether osteoporosis would be expected to negatively impact an MSC procedure can be made on both sides. For example, while MSC’s are clearly involved in the differentiation pathway that leads to osteoblasts and bone formation, they have been shown to be less osteogenic with advancing age in females [31-34]. On the other hand, animal models have demonstrated that thickening of bone trabeculae can be induced even with senile MSC’s [35].

It is important to clarify that the patients treated all presented with stable non-union fractures, meaning that excessive movement at the fracture line was absent. This inclusion criteria was used as one could postulate that unstable fractures would be less effective than stable non-union fractures without additional fixation and compression.
It is also interesting to note that no biologic scaffolds were used in any of these treatments. While biologic scaffolding combined with MSC’s and used for bone repair has become popular in the pre-clinical literature, these results would argue that they may not be necessary in some cases of non-union [16]. In fact, the use of a trocar or needle to cannulate the fracture site may well provide enough trauma to create a whole marrow scaffold. In this study, the placement of cells was accomplished via the use of a 22 gauge needle, which provides a less invasive approach than using a trocar, as described by Hernigou. In addition, Hernigou injected concentrated bone marrow mononucleated cells whereas in this current study, culture expanded MSC’s were used, this allows for a ready supply of purified stem cells that can be frozen for future use (i.e. a second application without a second harvest procedure). Finally, Koga has demonstrated that osteochondral repair may be initiated via direct application of MSC’s on the osteochondral defect without the use of scaffolding [36].

Many types of percutaneous therapy have been discussed to treat delayed unions and non-unions. However, many of these have significant side effects. For example, BMP’s have been shown to lead to nerve root irritation, ectopic bone formation, and antibody formation [37,38]. To the contrary, MSC’s have been shown to be neuroprotective to the dorsal root ganglia and other peripheral nerves in animal studies [39-42]. In addition, the use of PMMA cement has also been shown to result in bone formation in unwanted non-target tissues [43]. No ectopic bone formation was seen in this small case series and no complications were observed.

Platelet rich plasma (PRP) is defined as a concentrate of platelets re-suspended in plasma. PRP has been shown to assist fracture healing [44-46]. While not used here, autologous platelet lysate (PRP that has been lysed via freezing) was used in addition to MSC’s. We obviously cannot rule out that platelet derived growth factors may have also played a role in the observed effects here.

Conclusions

We report these results as preliminarily encouraging rather than conclusive. While this is a small case series, the fact that a combination of expanded bone marrow derived MSC’s and platelet derived growth factors can be delivered via trocar or needle to a fracture site may provide some advantages for the therapy of delayed and stable non-union fractures. In particular, such an approach is significantly less invasive than many surgical options including the placement of autologous bone grafts. While significant positive radiographic changes and concomitant increases in function were seen here in 4 of the 6 patients treated, larger, controlled studies need to be performed to further investigate the treatment approach.

References